



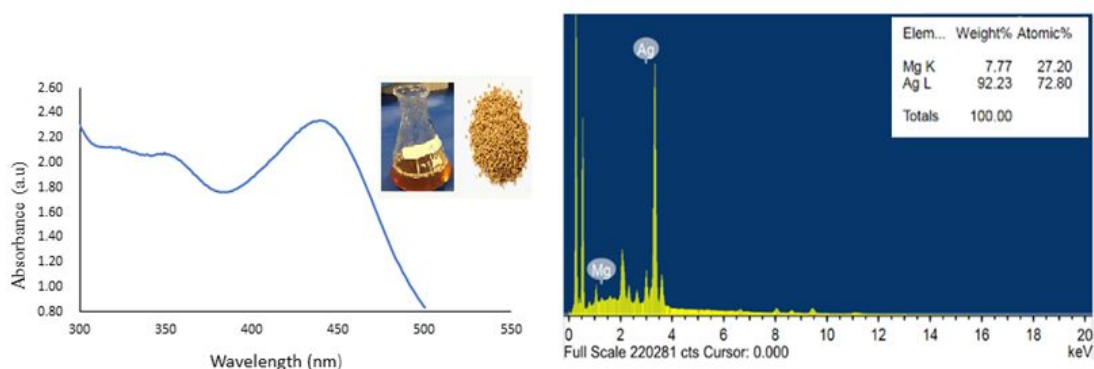
Synthesis and characterization of the photocatalytic and antibacterial properties of green silver nanoparticles

Journal:	<i>Green Chemistry Letters and Reviews</i>
Manuscript ID	TGCL-2019-0003.R2
Manuscript Type:	Research Letter
Date Submitted by the Author:	16-Aug-2019
Complete List of Authors:	Awad, Manal; 1King Abdullah Institute for Nanotechnology, Physics Hendi, Awatif; King Saud University, College of Science ALZahrani, Batool; King Saud University, College of Science Ortashi, khalid; King Saud University, College of engineering Soliman, Dina; King Saud University, College of Science Alanazi, Amnah; King Saud University, College of Science Alenazi, Wadha ; King Saud University, College of Science Mohammad Taha, Rasha ; Majmaah University, College of science Ramadan, Rasha ; King Saud university, fCentral Lab & Prince Naif Centre for Heath Science Research, King Saud University, Riyadh El-Tohamy, Maha; Knig saud University, Chemistry
Keywords:	Rhodamine B, Trigonella, Green synthesis, Catalysis, Antibacterial activity

SCHOLARONE™
Manuscripts

Graphical abstract:**Green synthesis and characterization of photocatalytic and antibacterial properties of silver nanoparticles**

Manal Ahmed Awad^{*a}, Awatif Ahnid Hendi^{*b}, Khalid Mustafa Ortashi^c, Batool Alzahrani^b, Dina Soliman^d, Amnah Alanazi^b, Wadha Alenazi^b, Rasha Mohammed Taha^c, Rasha Ramadan^f and Maha El-Tohamy^b

Green Synthesis of Silver nanoparticle using *Trigonella foenum*

Green synthesis and characterization of photocatalytic and antibacterial properties of silver nanoparticles

Manal Ahmed Awad^{*a}, Awatif Ahnid Hendi^{*b}, Batool Alzahrani^b, Khalid Mustafa Ortashi^c, Dina Soliman^d, Amnah Alanazi^b, Wadha Alenazi^b, Rasha Mohammed Taha^c, Rasha Ramadan^f and Maha El-Tohamy^b

^aKing Abdullah Institute for Nanotechnology, King Saud University, Riyadh 11451, Saudi Arabia. mawad@ksu.edu.sa

^bDepartment of Physics, King Saud University, Riyadh, 11495, Saudi Arabia. ahindi@ksu.edu.sa, amnahalanazi95@gmail.com, bt.00.l.ali@gmail.com, walenazi@KSU.EDU.SA

^cDepartment of Chemical Engineering, King Saud University, Riyadh, 11421, Saudi Arabia. ortashi9@ksu.edu.sa

^dDepartment of Microbiology, Faculty of Science, King Saud University, Riyadh, 11459, Saudi Arabia. dsoliman@ksu.edu.sa

^eDepartment of Physics , College of Science , Majmaah University, Majmaah 11932, Saudi Arabia. r.taha@mu.edu.sa

^fCentral Lab & Prince Naif Centre for Health Science Research, King Saud University, Riyadh, 11459, Saudi Arabia. rramadan@ksu.edu.sa

^gDepartment of Chemistry, King Saud University, Riyadh, 11495, Saudi Arabia star2000star@gmail.com

***Corresponding author:** Manal Ahmed Awad email: mawad@ksu.edu.sa & a.manalawad@gmail.com

Abstract

The present study focused on exploiting of eco-friendly *Trigonella foenum* extract for the synthesis of silver nanoparticles with minimum scale size and ultra-stable features.

The role of green synthesized nanoparticles was impact in improving and minimizing the environmental pollution by decreasing the use of hazardous chemicals and eliminating biological risks in biomedical applications. The as prepared silver nanoparticles were monitored by different spectroscopic and microscopic techniques. These include, UV-Vis spectrophotometry with maximum absorption peak at 443 nm, fourier transform infrared spectrophotometry and fluorescence analysis, transmission zetasizer, electron microscope and energy dispersive spectrometer. The morphology

of silver nanoparticles was predominantly spherical and aggregated into an irregular structure with an average diameter of 82.53 nm. Additionally, the prepared nanoparticles were employed for the screening of antibacterial activity and the photocatalytic degradation of hazardous *Rhodamine B* dye.

Keywords: *Rhodamine B*; *Trigonella foenum*; Green synthesis; Catalysis; Antibacterial activity

1. Introduction

Nanoscience and nanotechnology are the manipulation and the study of extremely small scale matter and can be employed in various applications across other science fields, such as physics, biology, chemical engineering and material science (1). Nanoparticles (NPs) are almost the final results of physicochemical and biological processes (2). The unique features of metal NPs such as, large surface energy, high surface area and quantum confinement, etc. might open expanding areas for researchers to exploit their potential applicability in various fields such as analytical chemistry (3), electronics (4), fuel cell (5) and biomedical applications (6). Thus, it is promising to use green and environmentally friendly methods for synthesis of NPs. Due to the presence of various natural chemical compounds such as polyphenols, flavonoids, sterols, triterpenes, and sugars in the plants, the literature survey revealed several green methods based on the employment of plant extracts (7, 8), or naturally capping materials (9) for the synthesis of metal NPs such as gold and silver NPs. The synthesized silver NPs (AgNPs) exhibited effective inhibitory activity against waterborne pathogens and microbes (10). Beside the valuable properties of AgNPs in various science fields such as optical, electronic, chemical, photoelectrochemical, catalytic, magnetic, they have significant antimicrobial activity and can be applied to the biomedical applications such as blood collection vessels, and wound bandages

1
2
3 (11). On the other hand, due to their cytotoxicity features, AgNPs have been reported
4 as nontoxic agents for animal cells, but highly toxic for bacterial cells such as
5
6 *Escherichia coli*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*. Therefore,
7
8 they can be act as effective bactericidal agents (12-13).
9
10

11
12 Organic dyes are considered as the major source of pollutants widely used in many
13 fields such as in industry of plastic, textile and medicine. The scientific researchers
14 started to concern with the hazardous influence of organic dyes as a major
15 environmental pollution problem (14). These industries required large quantities of
16 dyes that cause toxicity. Therefore, the necessary of new techniques to reduce their
17 accumulation and sever harmful effect is the main attention now.
18
19

20
21 Nowadays, the use of nanoparticles of nobel metals in photocatalytic degradation
22 provides many advantages over the conventional methods. Many researchers started
23 to study how to improve the photocatalytic degradation using nanometals such as
24 silver and gold (15, 16). The physical and chemical features of nanoparticles
25 including their reduced size, large surface area and high reactivity, play an important
26 role in the degradation of dyes (17). Therefore, the removal of non-biodegradable
27 dyes from the environment is considered as a crucial ecological problem (18).
28
29 Although, there are numerous available techniques to remove these dyes; recently,
30 metal NPs have been used to face this problem.
31
32

33
34 The present study aimed to suggest a new green synthesized AgNPs using *Trigonella*
35 *foenum* extract to degrade and illuminate the dye *Rhodamine B* (RB). Furthermore,
36 the antibacterial activity of the green synthesized AgNPs was screened against
37 different strains of bacteria.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

2. Materials and methods

2.1. Green synthesis of AgNPs

Trigonella foenum seeds were purchased from a local market in Riyadh City, Saudi Arabia. The seeds were initially washed in distilled water and dried on paper towels. Approximately, 10 g sample was grinded to powder and boiled in 100 mL distilled water for 30 min. The extract was filtered using a Whatman filter paper No.1 and the filtrate was kept at 40°C for further usage. Pure analytical grade of 2.0 mmol L⁻¹ silver nitrate (Techno Pharmchem, Delhi, India) was dissolved in 50 mL distilled water under vigorous stirring at 100 °C for 10 min. Then, approximately, 10 mL of *Trigonella* extract was added to the silver nitrate solution. The color of the colloidal solution was changed within ~30 min, which indicated the reduction of Ag ions into Ag metal and confirmed the formation of green AgNPs. The green AgNPs solution was centrifuged at 9000 rpm for 15 min and AgNPs pellets were collected, lyophilized and recovered in powdered using a Heto-Holten A/S, DK-3450 freeze dryer (Allerd, Denmark).

2.2. Characterization of AgNPs

The synthesized AgNPs were characterized using a Shimadzu UV 2450 UV-Vis Spectroscope (Shimadzu Corporation, Kyoto, Japan) over 300 –550 nm. The fluorescence and optical properties were characterized by a fluorescence spectrofluorophotometer (XRF, RF-5301(PC) S Shimadzu Corporation). Fourier Transform Infrared Spectroscopy (FTIR) for the prepared NPs was performed using Perkin Elmer FT-IR spectrophotometer (PerkinElmer Ltd., Yokohama, Japan). The size of the synthesized green AgNPs was analyzed using a dynamic light scattering (DLS) technique through a zetasizer (Nano series, HT Laser, ZEN3600 Malvern Instruments, Malvern, UK). The particle size, shape and morphology of the green

synthesized AgNPs were investigated using TEM (JEM-1011, JEOL Ltd., Tokyo, Japan) at an accelerating voltage of 100 kV. Energy-dispersive spectrometer (EDS) analysis was carried using a JEOL LTD JSM-7610F scanning electron microscope to confirm the presence of silver in the particles as well as to detect other elementary components of the particles.

2.3. Photocatalytic activity

In order to evaluate the photocatalytic activity of the green synthesized AgNPs, RB dye was degraded under light irradiation. A sample of ~ 5 mg of AgNPs was mixed with 20 mL of 10 mgL⁻¹ RB dye solution using a magnetic stirrer in the dark to maintain the equilibrium constant. After, exposing the solution to a UV lamp with constant stirring, the supernatant solution was measured using UV-Vis spectrophotometer.

Optical absorption spectra were determined for different durations of light exposure. A Perkin Elmer (Waltham, MA, USA) UV-Vis spectrophotometer was used to monitor the rate of degradation by recording the reduction in absorption intensity of dye at the maximum wavelength ($\lambda_{\max} = 552$ nm). The degradation efficiency (DE) was calculated by the following equation:

$$DE\% = \frac{A_0 - A}{A_0} \times 100$$

Where, A_0 is the initial absorption intensity at $\lambda_{\max} = 553$ nm and A is the absorption intensity after photocatalytic degradation.

2.4. Assessment of antibacterial activity

Antibacterial activity of synthesized green AgNPs was performed by using the agar well diffusion method (19, 20). Approximately 20 mL of sterile nutrient agar (Scharlau Microbiology, Barcelona, Spain) was poured into the sterile petri dishes. Triplicate plates were swabbed with overnight cultures of human pathogens *E. coli*

1
2
3 *ATCC 25922 S. aureus* ATCC 25923, and *Bacillus cereus* ATCC 11778 (clinical
4 isolates obtained from King Khalid University Hospital Riyadh, Saudi Arabia). The
5 solid medium was gently punched with a sterile cork borer to make a well
6 (3 wells/plate). Finally, the aqueous *Trigonella foenum* AgNPs (5, 15 $\mu\text{g mL}^{-1}$) were
7 added to each well and incubated for 24 h at 37°C. After complete incubation, the
8 diameter of the zone of inhibition was measured in cm.
9

16 3. Results and Discussion

17 3.1. UV-Vis spectral analysis of green synthesized AgNPs

18
19
20
21 UV-visible spectroscopy is a widely applied technique used to characterize and
22 determine the formation of nanoparticles and their optical properties. Due to the
23 influence of NPs spectrum by size, shape, interparticle interaction, free electron
24 density, it is considered as efficient tool for monitoring the electron transfer and
25 aggregation of NPs (21). The as prepared AgNPs using *Trigonella foenum* seeds
26 extract as reducing agent (22) was characterized using UV-Vis spectrophotometry. It
27 was observed that the initial pale yellow color was changed to dark brown due to the
28 excitation of the surface plasmon resonance (SPR) vibrations of the noble metal NPs,
29 indicating the reduction of Ag^+ in silver nitrate to the noble AgNPs. Figure 1 showed
30 the UV-Vis spectrum of AgNPs, with an absorption/extinction peak band at 443 nm.
31 The variation of intensity and bandwidth was resulted from the variation in size and
32 size distribution of the NPs. Furthermore, it was been found that there is an increase
33 in the integrated peak area of the band, which indicated a decrease in interparticle
34 spacing, due to the particle aggregation (23-25).
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52

53 3.2. Fluorescence studies of green synthesized AgNPs

54
55
56 The fluorescence properties of green synthesized AgNPs were investigated at λ_{ex} 450
57 nm and λ_{em} 662 nm. To the best of our knowledge, the surface activity of metal NPs
58
59
60

tends to react with hydroxide molecules could be enhanced by the large surface area per unit mass of metal NPs. Therefore, the hydroxyl groups of the extract exhibited a strong affinity towards the electrons. This phenomenon can be confirmed by the SPR of the AgNPs. It was observed that the SPR signal and the fluorescence were enhanced after the adsorption process (Figure 2). Furthermore, electromagnetic excitation induces electrons at the higher occupied energy level (HOEL) undergo electronic transitions to the lowest unoccupied energy level (LUEL) or excited states, the observed fluorescence was affected by the interaction between the **missive centers** and the interface environment. This can be explained that partial oxidation of metal NPs causes the formations of metal oxide clusters on the metal NP surface, which photoactivated by light excitations (26).

3.3. TEM and particle size distribution measurement

The green prepared AgNPs using *Trigonella foenum* **seed** extract was investigated microscopically using TEM. The morphology and size details of the formed AgNPs were provided and the TEM image depicted in Figure 3A, showed the appearance of spherical shaped AgNPs in individual form as well as aggregated particles.

Additionally, DLS is the most facile technique for evaluating the diffusion coefficient of particles in liquids. The average size distribution and hydrodynamic diameters of the nanoparticles can be calculated (27). Figure 3B, showed monodispersed AgNPs with an average diameter ~ 82 nm, with polydispersity index (PDI) of 0.134 revealing good colloidal dispersion and long term stability. The aggregation of AgNPs also estimated using PDI, values ranged from 0-1. Generally, values of ≤ 0.3 are considered to be acceptable and detect a homogenous distribution, and values > 0.7 indicate a heterogeneous dispersion of the samples and are usually not suitable to be measured by DLS (28).

3.4. EDS analysis of green synthesized AgNPs

Due to surface plasmon resonance, the metallic silver nanocrystals showed a typical optical absorption peak at the ~ 3 KeV (29, 30). Silver (92.23%) was the most common constituent element compared to magnesium (7.77%) as demonstrated in Figure 4. The EDS profile displayed a sharp signal for silver particles, which may be attributed to the biomolecules of *Trigonella foenum* extract bound to the surface of AgNPs, revealing their reduction of elemental silver.

3.5. FT-IR analysis of green AgNPs

The presence of biomolecules in the dispersed AgNPs was estimated using FT-IR. A comparative study between the FT-IR spectra of AgNPs (A) and *Trigonella foenum* seeds extract (B) was carried out and it was observed that synthesized AgNPs exhibited significant absorption bands at 3264.32, (2166.98 and 2144.50), 1636.98 and 1961.83 cm^{-1} were corresponding to -OH group, $-\text{C} \equiv \text{C}-$, N-H, and $=\text{C}-\text{H}$, respectively (Figures 5A and 5B). On the other hand, the previous FT-IR comparative spectra investigation showed a significant peak position shift, which may be attributed to the presence of the residual *Trigonella foenum* extract as reducing and stabilizing or capping agents for AgNPs (31). Furthermore, the presence of alkyne ($-\text{C} \equiv \text{C}-$) and amine ($-\text{NH}$) groups in *Trigonella foenum* extract were mainly involved in the reduction process of silver ions to AgNPs, which also confirmed that the presence of protein in *Trigonella foenum* extract could be acted as a stabilizing and reducing agents for AgNPs and preventing their agglomeration.

3.6. Photocatalytic degradation

1
2
3 The photocatalysis of the green synthesized AgNPs to induce RB degradation was
4 studied. It was observed that the photodegradation of RB dye was increased by
5 passing time and the complete photodegradation nearly (93%) with decolorization
6 was achieved after 216 h. Regarding to Beer's-Lambert law, a linear relationship was
7 obtained between the concentrations of RB vs. the intensity of the absorption peak at
8 443 nm. This can be attributed to the stable number of the photons available for
9 photodegradation. The layer of reducing agent on the surface of AgNPs may also
10 promote the effective adsorption between AgNPs and RB molecules. As a
11 consequence, the oxidation-reduction between the active RB and reducing agent can
12 occur more easily, effectively, and rapidly for particles of a smaller size (32). The
13 results indicated that the high reactivity of large surface area of AgNPs acted as an
14 efficient photocatalysis for dye degradation under UV light and confirmed the high
15 efficiency of AgNPs to minimize and eliminate the dye pollution. Owing to the
16 reaction mechanism is not fully known; oxidation-reduction reaction can be occurred
17 involving intermediates such as hydroxyl radicals (33).

37 3.7. Antibacterial activity of green AgNPs

38 The antibacterial activity of AgNPs synthesized by *Trigonella foenum* extract was
39 investigated using two concentrations $5 \mu\text{g mL}^{-1}$ (Ag 2) and $15 \mu\text{g mL}^{-1}$ (Ag 1) against
40 pathogenic organisms *S. aureus*, *B. cereus*, and *E. coli*. The well diffusion method
41 was applied and the diameters of the zone of inhibition /cm using AgNPs were shown
42 in Figure 7. The AgNPs synthesized by *Trigonella foenum* extract were found to have
43 higher antibacterial activity at both high and low concentrations against *E. coli* (1.4,
44 0.2 cm) and *B. cereus* (1, 0.09 cm), respectively, while less antibacterial activity for
45 the green AgNPs was found against *S. aureus* (0.5, 0.2 cm). Additionally, there was
46 no inhibition zone observed in the *Trigonella foenum* extract (Figure 7).
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Several studies have proposed the mechanisms underlying the bactericidal action of AgNPs, one of them suggested that, due to their high surface area, the AgNPs showed efficient antibacterial activity compared to the extract. This high surface area provides better contact with microorganisms. The NPs is attached to the cell membrane and penetrate the bacterial cell, through the interaction of AgNPs with sulfur-containing proteins contained in the bacterial membrane, as they do with phosphorus-containing compounds such as DNA. Another study suggested that when AgNPs penetrate the bacterial cell, they form a low molecular weight region in the center of the bacteria that conglomerates, protecting DNA from the silver ions. A third study suggested that, the NPs prevent cell division, which ultimately leading to cell death, by preferentially attack the respiratory electron transport chain. The NPs enhance their bactericidal activity, by release silver ions in the bacterial cells. Also, the action of AgNPs on bacteria may also occur via interaction with thiol group-containing respiratory enzymes of bacterial cells that inhibits the respiration process. In addition to that, it has been shown that AgNPs can enter into bacterial cells and hence condense DNA, thus preventing DNA from replicating and cells from reproducing (34-38).

Conclusion

The present study described a simple, rapid, and efficient method for the green synthesis of AgNPs using *Trigonella foenum* seed extract as a reducing agent. The optical properties, structure, morphology, photocatalytic activity and antibacterial activities of these AgNPs were examined. The optical analysis resulted in the appearance of a significant peak at 443 nm which confirmed the formation of AgNPs in the reaction mixture. FTIR spectrum identified the potential biomolecules in the *Trigonella foenum* extract which enhanced the reduced of Ag⁺ ions to Ag⁰. Spherical-shaped AgNPs with an average diameter of 82 nm were observed using

1
2
3 TEM, a zetasizer, and metal Ag signatures were confirmed by EDS. The antibacterial
4 activity of green synthesis AgNPs was exhibited to be effective against both Gram-
5 negative and Gram-positive strains. Finally, the obtained results revealed the
6 suitability of AgNPs as active material in the photocatalytic degradation of RB dye.
7
8
9

10 11 12 **Acknowledgments**

13
14 The authors are grateful for the financial and logistic support of King Abdullah
15 Institute for Nanotechnology and the Deanship of Scientific Research, King Saud
16 University, Riyadh, Saudi Arabia.
17
18
19

20 21 22 **Conflict of interest**

23
24 The authors declare that this work not associated with any conflict of interest
25

26 27 28 **Note of authors**

29
30 **Dr. Manal Ahmed Awad:** She received her M.Sc. and Ph.D. from Khartoum
31 University, Sudan in the field of physics. She has published more 20 scientific papers
32 all in international journals and patents. **Prof. Awatif Ahmed Hendi:** She received her
33 promotion to full Professor at King Saud University in 2010 in the field of physics.
34 She has published more 100 scientific papers all in international journals and more
35 than 10 patents. **Eng. Khalid Mustafa Ortashi:** He received his Master of Science
36 Water Resources Engineering from Free University Brussels, Belgium. He has
37 published more 20 scientific papers all in international journals and patents. **Ms.**
38 **Batool ALZahrani:** She is a postgraduate student under supervision of prof. Awatif
39 Ahmed Hendi (2017). **Researcher. Dina Soliman:** She has a master's degree (Plant
40 Science), college of science, Alazhar University, Egypt. **Ms. Amnah Alanazi:** She is a
41 postgraduate student under supervision of prof. Awatif Ahmed Hendi (2017).
42 **Researcher. Wadha Alenazi:** She has a master's degree (Physics), college of science,
43 King Saud University, Saudi Arabia. **Dr. Rasha Mohammad Taha:** She has a PhD
44 degree (Physics), college of science, Sudan University, Sudan. **Researcher Rasha**
45 **Ramadan:** She has a master's degree (biochemistry) college of science, King Saud
46 University. **Maha El-Tohamy:** is currently a Researcher in Department of Chemistry,
47 Faculty of Science, King Saud University. The author received her Ph.D.,
48 Pharmaceutical Sciences from Al-Azhar University, Cairo, Egypt in 2010, where her
49
50
51
52
53
54
55
56
57
58
59
60

analytical research was towards the green chemistry to minimize the formation of hazardous compounds.

References

1. Nouailhat, A. *An Introduction to Nanoscience and Nanotechnology*, Wiley Online Library, Hoboken, **2008**.
2. Aitken, R.J.; Creely, K.S.; Tran, C. *Nanoparticles: An occupational hygiene review*, HSE Research Report 274, London, **2004**.
3. Da Silva, B.F.; Barcelo, D. Analytical chemistry of metallic nanoparticles in natural environments. *TrAC Trends Anal. Chem.* **2011**, *30*, 528-540. doi:10.1016/j.trac.2011.01.008
4. Banu, K.; Harthi, K.; Gungune, M.S.; Gowri, V.P. Application of nanotechnology in medicine and electronics. *Int. J. Inn. Res. Comp. Commun. Eng.* **2017**, *5*, 138-140.
5. An, J.; Jeon, H.; Lee, J.; Chang, I.S. Bifunctional silver nanoparticle cathode in microbial fuel cells for microbial growth inhibition with comparable oxygen reduction reaction activity. *Env. Sci. Tech.* **2011**, *45*, 4551-5446. doi: 10.1021/es2000326
6. Alarfaj, N.A.; El-Tohamy, M.F.; Oraby, H. New label-free ultrasensitive electrochemical immunosensor-based Au/MoS₂/rGO nanocomposites for CA 27-29 breast cancer antigen detection. *New J. Chem.* **2018**, *42*, 11046-11053. doi: 10.1039/C8NJ01388H
7. Gabriela, A.M.; Gabriela, M.O.V.; Luis, A.M.; Reinaldo1, P.R.; Michael, H.M.; Rodolfo, G.P.; Roberto, V.B.J. Biosynthesis of silver nanoparticles using mint leaf extract (*Mentha piperita*) and their antibacterial activity. *Adv.Sci. Eng. Med.* **2017**, *9*, 1-10. doi:10.1166/asem.2017.2076

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
8. Ahmed, S.; Ahmed, S.M.; Swami, B.L.; Ikram S. Green synthesis of silver nanoparticles using *Azadirachta indica* aqueous leaf extract. *J. Rad. Res. Appl. Sci.* **2016**, *9*(1), 1-7. <https://doi.org/10.1016/j.jrras.2015.06.006>
 9. Luo, L.J.; Lin, T.Y.; Yao, C.H.; Kuo, P.Y.; Matsusaki, M.; Harroun, S.G.; Huang, C.C.; Lai, J.Y.; Dual-functional gelatin-capped silver nanoparticles for antibacterial and antiangiogenic treatment of bacterial keratitis. *J Coll. Inter. Sci.* **2019**, *536*, 112-126. <https://doi.org/10.1016/j.jcis.2018.10.041>
 10. Aswathy Aromal, S.A.; Philip, D. Green synthesis of gold nanoparticles using *Trigonella foenum-graecum* and its size-dependent catalytic activity. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2012**, *97*, 1-5. doi: [10.1016/j.saa.2012.05.083](https://doi.org/10.1016/j.saa.2012.05.083)
 11. Sreeprasad, T.S.; Pradeep, T. *Noble Metal Nanoparticles*, in: R. Vajtai (Ed.), Springer Handbook of Nanomaterials, Springer-Verlag, Berlin, **2013**, pp. 303–388.
 12. Awad, M.; El Dib, R.; Almusayeib, N.; Massarani, S.; Ortashi K.M.; Hendi, A. Novel balanites aegyptiaca mesocarp synthesized silver nanoparticles: formation, characterization, antimicrobial, cytotoxicity and antiviral effects. *Dig. J. Nanomater. Bios.* **2013**, *8*(4), 1665-1677.
 13. Logeswari, P.; Silambarasan, S.; Abraham, J. Synthesis of silver nanoparticles using plants extract and analysis of their antimicrobial property. *J. Saudi Chem. Soc.* **2015**, *19*(3), 311-317. <https://doi.org/10.1016/j.jscs.2012.04.007>
 14. Faisal, M.; Tariq, M.A.; Muneer, M. Photocatalysed degradation of two selected dyes in UV-irradiated aqueous suspensions of titania. *Dyes Pigm.* **2007**, *72*, 233–239. <https://doi.org/10.1016/j.dyepig.2005.08.020>
 15. Arunachalam, R.; Dhanasingh, S.; Kalimuthu, B.; Uthirappan, M.; Rose, C.; Mandal, A.B. Phytosynthesis of silver nanoparticles using *Cocciniagrandsis* leaf

- 1
2
3 extract and its application in the photocatalytic degradation. *Colloids Surf. B*
4 *Biointerfaces*, 2012, 94, 226-230. doi: 10.1016/j.colsurfb.2012.01.040.
5
6
7
8 16. Suwith, V.S.; Philip, D. Catalytic degradation of methylene blue using
9 biosynthesized gold and silver nanoparticles. *Spectrochim. Acta A Mol. Biomol.*
10 *Spectrosc.* 2014, 118, 526-532. <https://doi.org/10.1016/j.saa.2013.09.016>
11
12
13
14 17. Ghosh, S.K.; Kundu, S.; Mandal, M.; Pal, T. Silver and gold nanocluster
15 catalyzed reduction of methylene blue by arsine in a micellar medium. *Langmuir*
16 *2002*, 18, 8756–8760. <https://doi.org/10.1021/la0201974>
17
18
19
20
21 18. Bogireddy, N.K.R. Kumar, H.A.K. Mandal, B.K. Biofabricated
22 silver nanoparticles as green catalyst in the degradation of
23 different textile dyes. *J. Environ. Chem. Eng.* **2016**, 4(1), 56-64.
24 doi: 10.1016/j.jece.2015.11.004
25
26
27
28
29
30
31 19. Magaldi, S.; Mata-Essayag, S.; Hartung de Capriles, C.; Perez, C.; Colella, M.T.;
32 Olaizola, C.; Ontiveros, Y. Well diffusion for antifungal susceptibility testing. *Int.*
33 *J. Inf. Diseases* **2004**, 8(1), 39-45. doi:10.1016/j.ijid.2003.03.002
34
35
36
37
38 20. Valgas, C. Souza, S.M.D.; Smânia, E.F.A, Smânia Jr., A. Screening methods to
39 determine antibacterial activity of natural products. *Braz. J. Microbiol.* **2007**, 38,
40 369-380. <http://dx.doi.org/10.1590/S1517-83822007000200034>
41
42
43
44 21. Rucha, D.; Mankad, V.; Gupta, S.; Jha, P. Size Distribution of Silver
45 Naoparticles: UV-Visible Spectroscopic Assessment *Nanosci. Nanotech. Lett.*
46 **2012**, 4, 30-34. doi: 10.1142/S0219581X12500135
47
48
49
50
51 22. Varghese, R.; Almalki, M.A.; Ilavenil, S; Rebecca, J.; Choi, K.C.; Silver
52 nanopaticles synthesized using the seed extract of *Trigonella foenum graecum* L.
53 and their antimicrobial mechanism and anticancer properties *Saudi. J. Biol. Sci.*
54 **2017**, doi: <http://dx.doi.org/10.1016/j.sjbs.2017.07.001>
55
56
57
58
59
60

- 1
2
3 23. Addison, C.J.; Brolo, A.G. Nanoparticle-containing structures as a substrate for
4 surface enhanced raman scattering. *Langmuir*, **2006**, *22*(21), 8696-8702.
5
6 [doi: 10.1021/la061598c](https://doi.org/10.1021/la061598c)
7
8
9
10 24. Meyer, M.; Le, R. E.C.; Etchegoin, P.G. Self limiting aggregation leads to long
11 lived metastable clusters in colloidal solutions. *J. Phys. Chem. B*, **2006**, *110*(12),
12 6040-6047. [doi: 10.1021/jp055866b](https://doi.org/10.1021/jp055866b)
13
14
15
16 25. Seney, C.S.; Gutzman, B.M.; Goddard, R.H. Correlation of size and surface
17 enhanced raman scattering activity of optical and spectroscopic properties
18 for silver nanoparticles. *J. Phys. Chem. C*, **2009**, *113*(1), 74-80.
19
20
21
22 [doi: 10.1021/la061598c](https://doi.org/10.1021/la061598c)
23
24
25
26 26. Peyser, L.A.; Vinson, A.E.; Bartko, A.P.; Dickson, R.M. Photoactivated
27 fluorescence from individual silver nanoclusters. *Sci*, **2001**, *291*(5501), 103-106.
28
29
30
31 [doi: 10.1126/science.291.5501.103](https://doi.org/10.1126/science.291.5501.103).
32
33 27. Kaviya, S.; Santhanalakshmi, J.; Viswanathan, B.; Muthumary, J.; Srinivasan,
34 K. Biosynthesis of silver nanoparticles using citrus sinensis peel extract and its
35 antibacterial activity. *Acta A Mol. Biomol. Spectrosc.* **2011**, *79*(3), 594-598.
36
37
38
39 [doi: 10.1016/j.saa.2011.03.040](https://doi.org/10.1016/j.saa.2011.03.040).
40
41
42 28. Baalousha, M.; Lead, J.R. Rationalizing nanomaterial sizes measured by atomic
43 force microscopy, flow field-flow fractionation, and dynamic light scattering:
44 sample preparation, polydispersity, and particle structure. *Environ. Sci. Technol.*
45
46
47
48 **2012**, *46*(11), 6134-6142. <https://doi.org/10.1021/es301167x>
49
50
51 29. Jyoti, K.; Baunthiyal, M.; Singh, A. Characterization of silver nanoparticles
52 synthesized using *Urtica dioica* Linn. leaves and their synergistic effects with
53 antibiotics. *J. Radiat. Res. Appl. Sci.* **2016**, *9*(3), 217-227.
54
55
56
57
58
59
60

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
30. Danaei, M.; Dehghankhold, M.; Ataei, S.; Hasanzadeh, D. F.; Javanmard, R.; Dokhani, A.; Khorasani, S.; Mozafari, R.M. Impact of particle size and polydispersity index on the clinical applications of lipidic nanocarrier systems, *Pharmaceutics*, **2018**, *10*(2), 57.
31. Vanaja, M.; Paulkumar, K.; Baburaja, M.; Rajeshkumar, S.; Gnanajobitha, G.; Malarkodi, C.; Sivakavinesan M.; Annadurai, G. Degradation of methylene blue using biologically synthesized silver nanoparticles. *Bioinorg. Chem. App.* **2014**, *2014*, 1-8. <http://dx.doi.org/10.1155/2014/742346>
32. Vidhu, V.K.; Philip, D. Catalytic degradation of organic dyes using biosynthesized silver nanoparticles, *Micron*. **2014**, *56*, 54-62.
33. Peral, J.; Trillas, M.; Domenech, X. Heterogeneous Photochemistry: An Easy Experiment. *J. Chem. Educ.* **1995**, *72*(6), 565.
34. Sondi, B.; Salopek, S. Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *J. Colloid. Interface Sci.* **2004**, *275*(1), 177-182.
35. Jose, R. M.; Jose, L. E.; Alejandra, C.; Katherine, H.; Juan, B.K.; Jose, T. R.; Miguel, J. Y. The bactericidal effect of silver nanoparticles. *Nanotechnology*. **2005**, *16*(10), 2346.
36. Kvitek, L.; Panacek, A.; Soukupova, J.; Kolar, M.; Vecerova, R.; Pucek, R.; Holecova, M.; Zboril, R. Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). *J. Phys. Chem. C.* **2008**, *112*(15), 5825-5834.
37. Klasen, H. J. Historical review of the use of silver in the treatment of burns. I. Early uses. *Burns*. **2000**, *26*(2), 117-130.

- 1
2
3 38. Li, W.R.; Xie, X.B.; Shi, Q.S.; Duan, S.S.; Yang, Y. S.O.; Chen, Y. B
4
5 Antibacterial effect of silver nanoparticles on *Staphylococcus aureus*. *Biometals*.
6
7
8 **2011**, 24, 135–141.
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For Peer Review Only

List of Figure Captions

Figure 1. UV-Vis spectra of *Trigonella foenum* AgNPs

Figure 2. Fluorescence spectra of *Trigonella foenum* AgNPs

Figure 3. (A) TEM micrograph of *Trigonella foenum* AgNPs, (B) Zetasizer measurement of the average size of *Trigonella foenum* AgNPs

Figure 4. EDS spectrum showing a higher percentage of silver signals

Figure 5. FTIR spectrum of (A) *Trigonella foenum* AgNPs (B) *Trigonella foenum* seeds extract

Figure 6. Percentage of dye degradation at time of exposure

Figure 7. Antibacterial activity of two concentrations 15 $\mu\text{g mL}^{-1}$ and 5 $\mu\text{g mL}^{-1}$ of AgNPs synthesized using *Trigonella foenum* seeds extract against three bacterial strains: A) *E. coli*, B) *B. cereus*, C) *S. aureus*

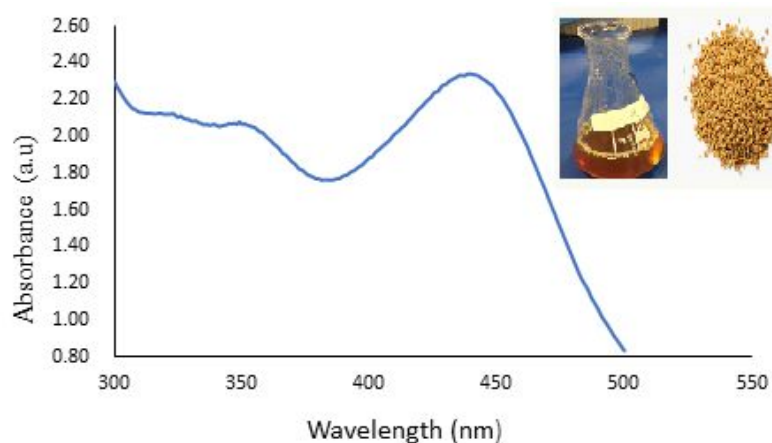


Figure 1. UV-Vis spectra of *Trigonella foenum* AgNPs

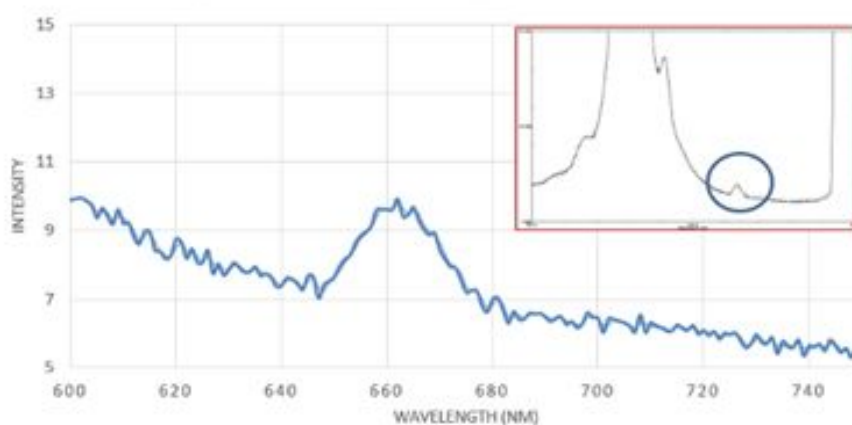


Figure 2. Fluorescence spectra of *Trigonella foenum* AgNPs

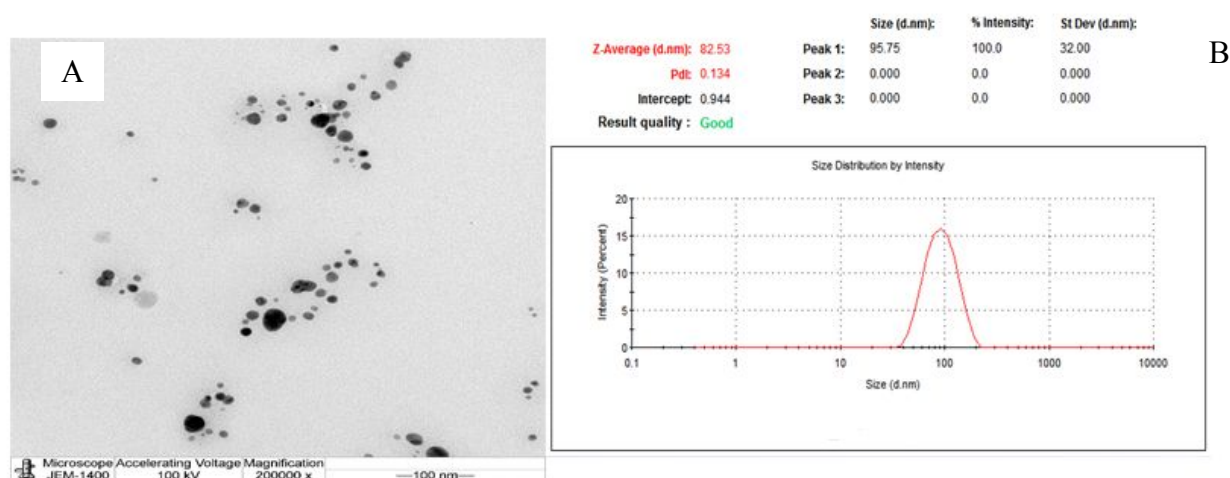


Figure 3. (A) TEM micrograph of *Trigonella foenum* AgNPs, (B) Zetasizer measurement of the average size of *Trigonella foenum* AgNPs

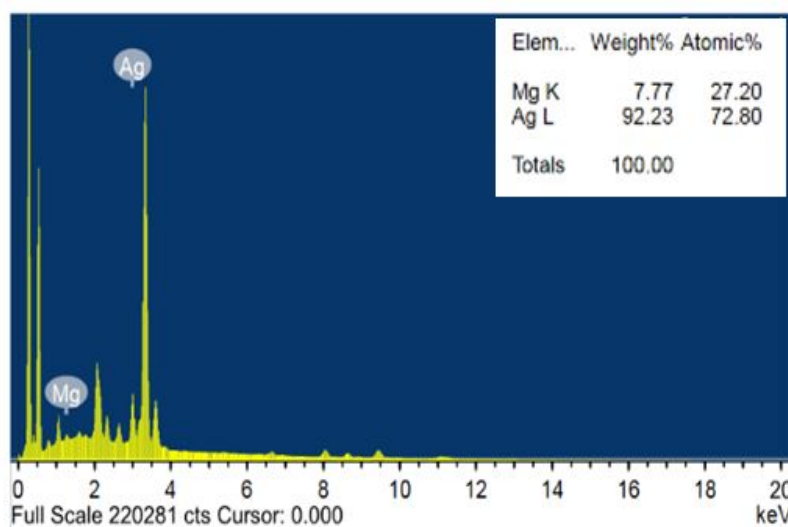


Figure 4. EDS spectrum showing a higher percentage of silver signal

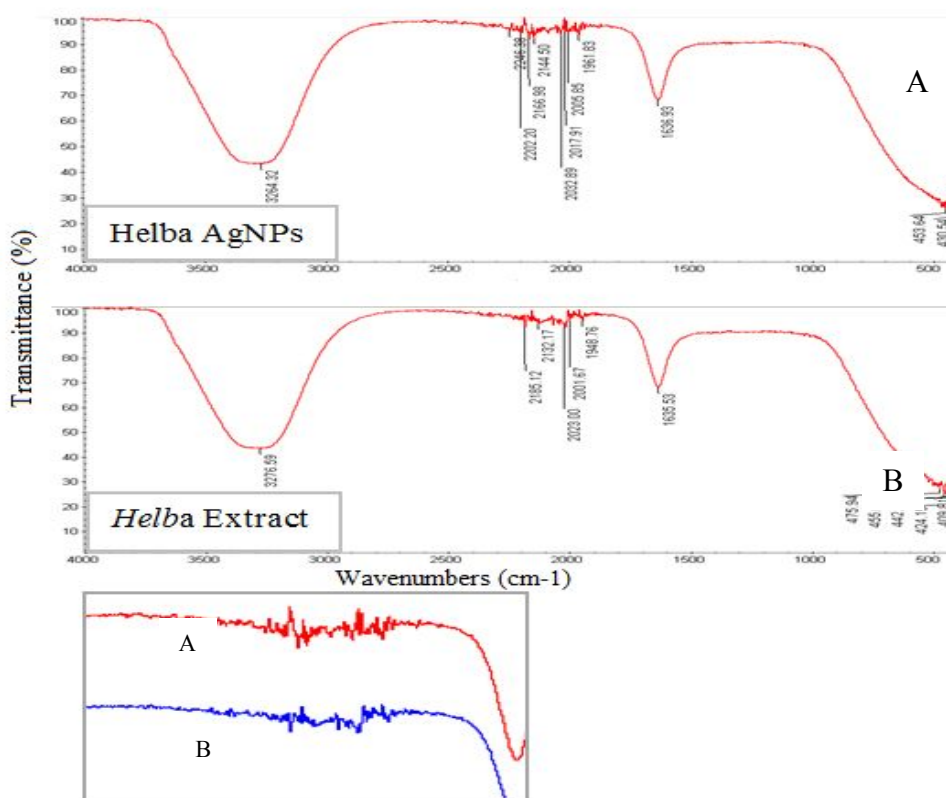


Figure 5. FTIR spectrum of (A) *Trigonella foenum* AgNPs (B) *Trigonella foenum* seeds extract

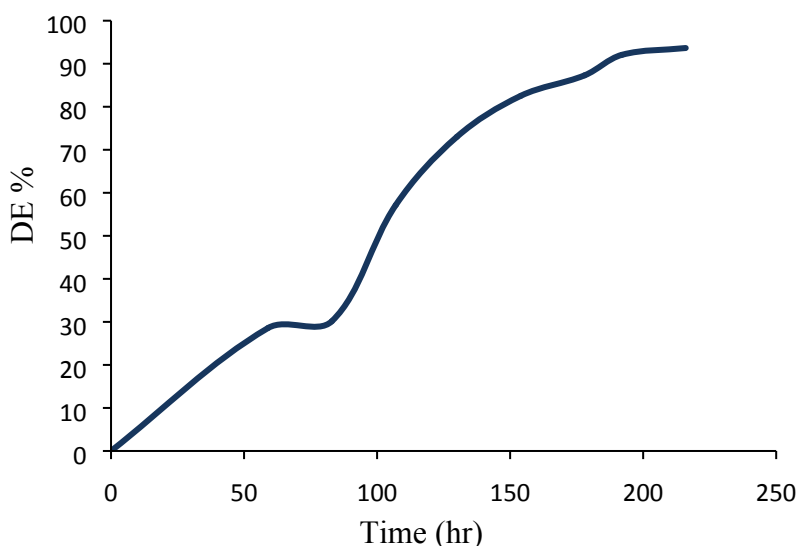


Figure 6. Percentage of dye degradation at time of exposure

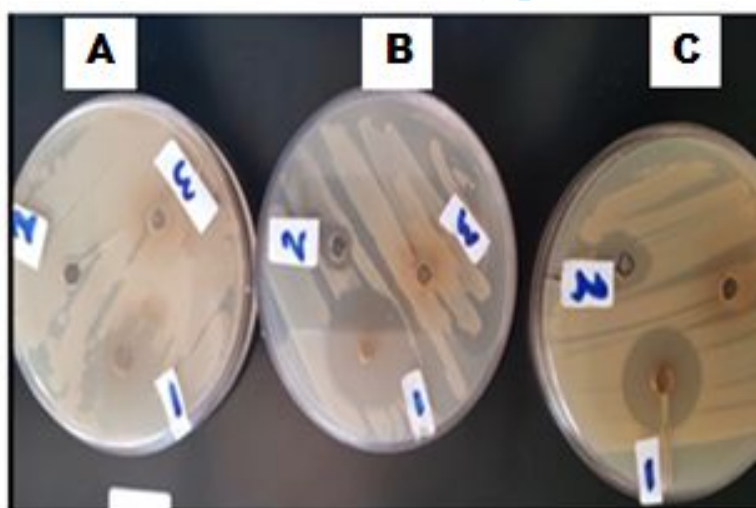
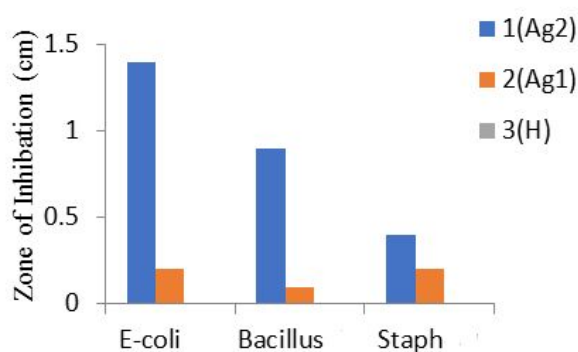


Figure 7. Antibacterial activity of two concentrations $15 \mu\text{g mL}^{-1}$ and $5 \mu\text{g mL}^{-1}$ of AgNPs synthesized using *Trigonella foenum* seeds extract against three bacterial strains: A) *E. coli*, B) *B. cereus*, C) *S. aureus*