Research Article

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Bactericidal and cytotoxic properties of green synthesized nanosilver using Rosmarinus officinalis leaves

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Abstract: Green synthesized nanoparticles from plant extracts are being used in various biomedical applications, particularly because of their bactericidal and cytotoxic activities. In this study, silver nanoparticles (AgNPs) were successfully synthesized from the Rosmarinus officinalis aqueous leaf extract. Different spectroscopic and microscopic analyses such as ultraviolet-visible (UV-vis) spectroscopy, Fourier transform infrared spectroscopy (FTIR), scanning electron microscopy, and energy-dispersive X-ray spectroscopy were performed to verify the biosynthesized AgNPs in our sample. The formation of nanosilver particles was preliminarily confirmed by UV-vis spectroscopy at 400 nm. The presence of carboxylic or amide groups was confirmed by FTIR, for the reduction of the silver ion. Transmission electron microscopy confirmed a particle size of 12-22 nm. The prepared AgNPs showed good antibacterial activity against human pathogens and good cytotoxic activity against the human breast cancer cell line (MDA MB 231). The nanoparticles prepared from R. officinalis can be used for various biomedical applications.

Keywords: Rosmarinus officinalis, green synthesis, nanosilver

List of abbreviations

AgNPs	Silver nanoparticles
DLS	Dynamic light scattering
EDX	energy-dispersive X-ray spectrum
FTIR	Fourier transform infrared spectroscopy
PDI	Polydispersity index
SPR	Surface plasmon resonance
TEM	Transmission electron microscopy
UV-vis	Ultraviolet-visible absorption

1 Introduction

Silver has been used by mankind for a long time because of its low toxicity and good biocidal properties. It is frequently used in water-purifying systems in hospitals, for healing injuries, and in the field of nanoparticle synthesis [1]. Some bacteria are resistant to silver and are able to accumulate this metal up to 25% of their dry weight biomass in their cells [2], which makes them suitable for extracting silver from ores [3]. Silver nanoparticles (AgNPs) are used as an effective nanodrug against many diseases [4]. Also, they are more efficient in killing silver-resistant bacteria due to their highly developed surface area [5], whereas highly reactive silver ions (Ag⁺ ions) form insoluble precipitates so are less efficient in killing bacteria [6].

Nanobiotechnology is constantly exploring the synthesis of metal nanoparticles in a nontoxic and ecofriendly manner [7]. The main requirements for the synthesis of a metal nanoparticle are the solvent medium, a reducing agent, and a nontoxic stabilizer of nanoparticles [8]. Plants are rich in active compounds and, thus, can be used as a good source of reducing agents for synthesizing nanoparticles [9]; this has

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prompted a new field of green synthesis [10]. Almost all parts of the plants are rich in active biomolecules such as amino acids, alkaloids, proteins, polysaccharides, phenolics, terpenoids, and flavones, which can exhibit good reducing and capping properties useful in synthesizing nanoparticles [11]. Additionally, hydroxyl and carboxyl groups present in phenols and flavonoids wrap the nanoparticles and act as capping agents [12].

Rosmarinus officinalis, commonly known as rosemary, contains many different bioactive compounds, and it is used as a flavoring spice in cooking for many years [13]. It is one of the most important commercial crops in Saudi Arabia [14]. Its essential oil has been reported to have analgesic, antispasmodic, antibacterial, and hypnotic effects [15]. It is also used in the treatment of depression and stress-related disorders [16]. Same plants grown under different climatic



Figure 1: UV-vis spectral analysis of the biosynthesized AgNPs prepared with aqueous *R. officinalis* leaf extract.

conditions show different active components [17,18]. Some environmental factors like type of soil and temperature can alter the plant structure and its phytochemical activity, especially the bioactive compounds present in the leaves [19]. Some studies even reported a direct relationship between vitamins and phenolic compounds of plant and soil mineral content of the area where the plant is grown [20]. Natural compounds of R. officinalis leaves change significantly if grown under differential environmental conditions [21]. It was of great interest to synthesize AgNPs by the reduction of aqueous Ag⁺ ions with the *R*. officinalis leaf extract collected from Rivadh, Saudi Arabia, and synthesis of AgNPs from R. officinalis collected from deserts has not been reported. The green synthesized AgNPs were characterized using various microscopic and analytical techniques, including absorption (UV-vis) spectroscopy, ultraviolet-visible Fourier transform infrared spectroscopy, and transmission electron microscopy (TEM). The bactericidal and cytotoxic properties of the synthesized nanoparticles were also analyzed.

2 Materials and methods

2.1 Leaf extract

Fresh green leaves of rosemary cultivated in the Riyadh region of Saudi Arabia were used for this study.



Figure 2: IR spectra of the biosynthesized AgNPs prepared with aqueous R. officinalis leaf extract.

Ten grams of healthy leaves were washed, chopped, and boiled in 0.1 L of double-distilled water for 30 min. After cooling to room temperature, the broth was filtered and stored at 4°C.

2.2 Biosynthesis of AgNPs

The freshly prepared leaf extract was mixed with a silver nitrate solution at a final concentration of 5 mM at room temperature. The reduction of silver ions to AgNPs was indicated by a color change from yellow to brown and recorded by scanning through a UV-vis spectrophotometer at different intervals for 3 h.

2.3 Characterization of the prepared nanoparticles

The as-prepared nanoparticles were analyzed by observing the color change of the reaction mixture. The absorbance of the reaction mixture was measured over the range of 200–700 nm every 30 min. The average size of the as-prepared nanoparticles was determined by carrying out the zeta potential analysis. The hydrodynamic size and zeta potential of the prepared AgNPs were measured at the scattering angle of 173° with a red laser at the wavelength of 633 nm. The average size of the synthesized particles was measured with the help of a TEM. Infrared spectroscopy was used to determine the nature of bio-reducing functional groups.



Figure 3: Average size of the biosynthesized AgNPs prepared with aqueous R. officinalis leaf extract.



Figure 4: Zeta potential distribution of the biosynthesized AgNPs prepared with aqueous R. officinalis leaf extract.

2.4 Antimicrobial activity

Bacterial strains, namely *Bacillus subtilis, Escherichia coli, Klebsiella pneumoniae, Enterococcus Faecalis, Salmonella typhi, Staphylococcus aureus, Staphylococcus epidermidis,* and *Streptococcus pneumoniae,* were provided by King Khalid Hospital, Riyadh, Saudi Arabia. The antibacterial activity of the synthesized AgNPs against all the mentioned bacteria was determined by the disk diffusion method.

2.5 Screening of cytotoxic activity of the prepared nanoparticles

R. officinalis nanoparticles were screened for cytotoxicity on the breast cancer cell line MDA-231 by the MTT assay. Different concentrations of the synthesized nanoparticles (3.125, 6.25, 12.5, 25, 50, and 100 μ L) were used to treat the cancerous cells. The inhibition percentage was calculated.

3 Results

UV-vis spectroscopy was used as a primary method to confirm the synthesis of nanoparticles due to their interaction with specific wavelengths. The synthesis of nanoparticles from silver ions in the presence of the rosemary plant extract was initially confirmed by a color change from yellow to brown. The surface plasmon resonance (SPR) peak at 400 nm confirmed the presence of nanoparticles in the extract (Figure 1). Free electrons in the nanoparticles resonate at certain wavelengths, resulting in SPR. A size reduction of the particles can lower the SPR peak, thus relating the absorption to the particle size [22].

Recorded infrared (IR) spectra point toward the active compounds of the *R. officinalis* extract, which facilitate the green synthesis of nanoparticles. Nanoparticles are synthesized in three phases, starting with the activation phase, where the plant metabolites with reducing capacities help to split the metal ion from the salt. Next is the growth phase, where metal ions combine to form nanoparticles, and finally, the termination phase, where capping around the synthesized particle is done with the help of the metabolites present in the plant extract [23]. The IR spectrum of Figure 2 shows an intense peak at 1636.75 cm⁻¹, which is recognized as C=O and C=C bond stretches and identified as carboxylic or amide groups, and a peak at 3304.40 cm^{-1} , which represents the

hydroxyl group and suggests the presence of phenols, flavonoids, and alcohols in the plant extract.

In this study, we used dynamic light scattering (DLS) to determine the particle size distribution of the synthesized AgNPs (Figure 3). The *Z*-average mean of the synthesized AgNPs was 66.79 nm, with a polydispersity index (PDI) of 0.177. The PDI is best measured with DLS. Our results with respect to nanoparticles size are acceptable, as a PDI value of more than 0.7 indicates a broad size distribution [24]. Plant extracts are good reducing agents for the synthesis of metal nanoparticles. The interface of the particles mainly depends on the zeta



Quantitative results



Figure 5: EDX elemental analysis of the biosynthesized AgNPs prepared with aqueous *R. officinalis* leaf extract.

potential since a larger zeta potential results in a particle abundance, thus providing stability to the particles (Figure 4) [25]. The energy-dispersive X-ray spectrum (EDX) of the synthesized nanoparticles exhibited strong signals in the oxygen, potassium, and chlorine regions (Figure 5). The presence of other elements such as carbon, calcium, sodium, and magnesium was also observed in the spectrum. A TEM image was used to find the average size of the prepared nanoparticles, which was in the range of 12–22 nm (Figure 6). TEM is typically an ultrathin sample image showing the 2D structure with the help of an absorption beam passing through the sample [26]. Nowadays, AgNPs are highly important in detergent and disinfectant manufacturing industries because of the resistance level of microbes to chemical fungicides and disinfectants [27]. We examined the antibacterial efficiency of AgNPs prepared from R. officinalis against some pathogenic bacteria. The varying degree of inhibition by both the Gram-positive and Gram-negative bacteria is shown in Table 1.

4 Discussion

The antibacterial efficacy of a particle depends on its small size and round shape. Our results agree with many previous studies reporting high antimicrobial activities **Table 1:** Antibacterial activity of the biosynthesized AgNPs prepared with aqueous *R. officinalis* leaf extract against eight bacterial species tested by disc diffusion assay

	Strains	Zone of inhibition (mm)
Gram positive	S. aureus	11 ± 0.30
	Staphylococcus epidermidis	8 ± 0.13
	S. pneumonia	10 ± 0.25
	Enterococcus Faecalis	$10\ \pm\ 0.50$
	B. subtilis	11 ± 0.20
Gram negative	E. coli	13 ± 0.10
	S. typhi	$14~\pm~0.31$
	K. pneumoniae	$9~\pm~0.11$

of round-shaped nanoparticles with a size of less than 10 nm [28]. It has been suggested that the antibacterial properties of the plant-derived AgNPs could be due to either the particles' ability to permeate the cell and interact with the genetic material deoxyribonucleic acid (DNA) and other important constituents, leading to cell death, or their adherence to the negatively charged cell surface that alters the chemical and physical properties of the cell wall and cell membrane [29]. In this study, the green synthesized AgNPs from *R. officinalis* were evaluated at various concentrations for the cytotoxic effects on the breast cancer cell lines (MB 231) *in vitro*. As observed in Figure 7, both plant extracts and nanoparticles decrease the viability of cancer cell lines (MB 231),



Figure 6: TEM micrograph of the biosynthesized AgNPs prepared with aqueous *R. officinalis* leaf extract (particle size shown in range from 12 to 22 nm).

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Figure 7: Cytotoxicity of aqueous *R. officinalis* leaf extract and its AgNPs on breast cancer cell lines (MDA MB 231) following 24 h exposure.

but the cytotoxicity of AgNPs was higher than the cytotoxicity of the plant extract. The cytotoxic effect of nanoparticles on cell viability has a major role in antitumor activity, thereby reducing disease progression [30]. The cytotoxic effects of silver result from the active physiochemical interaction of silver atoms with functional groups of intracellular proteins, as well as with the nitrogen bases and phosphate groups in DNA [31].

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References

- [1] Graham C. The role of silver in wound healing. Br J Nurs. 2005;14(19):S22.
- [2] Seifelnassr AAS, Abouzeid AZM. Exploitation of bacterial activities in mineral industry and environmental preservation: an overview. J Min. 2013;2013:507168.
- [3] Pooley FD. Bacteria accumulate silver during leaching of sulphide ore minerals. Nature. 1982;296(5858):642-3.
- [4] Rasheed T, Bilal M, Li C, Iqbal HMN. Biomedical potentialities of taraxacum officinale-based nanoparticles biosynthesized using methanolic leaf extract. Curr Pharm Biotechnol. 2017;18(14):1116–23.
- [5] Ying JY. The era of nanotechnology. Nano Today. 2008;3:1.
- [6] Sun Z, Fan C, Tang X, Zhao J, Song Y, Shao Z, et al. Characterization and antibacterial properties of porous fibers containing silver ions. Appl Surf Sci. 2016;387:828–38.
- [7] Rasheed T, Bilal M, Li C, Nabeel F, Khalid M, Iqbal HMN. Catalytic potentialof bio-synthesized silver nanoparticles using Convolvulus arvensis extract for the degradation of environmental pollutants. J Photochem Photobiol B. 2018;181:44–52.

- [8] Raveendran P, Fu J, Wallen SL. Completely "Green" synthesis and stabilization of metal nanoparticles. J Am Chem Soc. 2003;125(46):13940-1.
- [9] Barkat MA, Harshita, Beg S, Naim MJ, Pottoo FH, Singh SP, et al. Current progress in synthesis, characterization and applications of silver nanoparticles: precepts and prospects. Recent Pat Antiinfect Drug Discov. 2018;13(1):53–69.
- [10] Rasheed T, Bilal M, Iqbal HMN, Li C. Green biosynthesis of silver nanoparticles using leaves extract of Artemisia vulgaris and their potential biomedical applications. Colloids Surf B. 2017;1(158):408–15.
- [11] Ahumada M, Suuronen EJ, Alarcon EI. Biomolecule silver nanoparticle-based materials for biomedical applications.
 In: Martínez L, Kharissova O, Kharisov B, editors. Handbook of Ecomaterials. Cham: Springer. 2019.
- [12] Makarov VV, Love AJ, Sinitsyna OV, Makarova SS, Yaminsky IV, Taliansky ME, et al. "Green" nanotechnologies: synthesis of metal nanoparticles using plants. Acta naturae. 2014;6(1):35–44.
- [13] Borrás-Linares I, Stojanović Z, Quirantes-Piné R, Arráez-Román D, Švarc-Gajić J, Fernández-Gutiérrez A, et al. *Rosmarinus officinalis* leaves as a natural source of bioactive compounds. Int J Mol Sci. 2014;15(11):20585–606. doi: 10.3390/ijms151120585.
- [14] Bazaid SA, El-Amoudi MS, Ali EF, Abdel-Hameed ES. Volatile oil studies of some aromatic plants in Taif region. J Medicinal Plants Stud. 2013;1(5):119–28.
- [15] de Oliveira JR, Camargo SEA, de Oliveira LD. Rosmarinus officinalis L. (rosemary) as therapeutic and prophylactic agent. J Biomed Sci. 2019;26(1):5.
- [16] Fernández LF, Palomino OM, Frutos G. Effectiveness of *Rosmarinus officinalis* essential oil as antihypotensive agent in primary hypotensive patients and its influence on healthrelated quality of life. J Ethnopharmacol. 2014;151:509–16.
- [17] Guo H, Chamberlain SA, Elhaik E, Jalli I, Lynes AR, Marczak L, et al. Geographic variation in plant community structure of salt marshes: species, functional and phylogenetic perspectives. PLoS One. 2015;10(5):e0127781.
- [18] Liu W, Liu J, Yin D, Zhao X. Influence of ecological factors on the production of active substances in the anti-cancer plant sinopodophyllum hexandrum (Royle) T. S. Ying. PLoS One. 2015;10(4):e0122981.
- [19] Liu W, Yin D, Li N, Hou X, Wang D, Li D, et al. Influence of environmental factors on the active substance production and antioxidant activity in Potentilla fruticosa L. and its quality assessment. Sci Rep. 2016;6:28591.
- [20] Tsao R, Khanizadeh S, Dale A. Designer fruits and vegetables with enriched phytochemicals for human health. Can J Plant Sci. 2006;86:773–86.
- [21] Peñuelas J, Llusià J. Effects of carbon dioxide, water supply, and seasonality on terpene content and emission by *Rosmarinus officinalis*. J Chem Ecol. 1997;23:979.
- [22] Peng S, McMahon JM, Schatz GC, Gray SK, Sun Y. Reversing the size-dependence of surface plasmon resonances. Proc Natl Acad Sci U S A. 2010;107(33):14530-4.
- [23] Malik P, Shankar R, Malik V, Sharma N, Mukherjee T. Green chemistry based benign routes for nanoparticle synthesis. J Nanopart. 2014;2014:302429.
- [24] Clayton KN, Salameh JW, Wereley ST, Kinzer-Ursem TL. Physical characterization of nanoparticle size and surface

modification using particle scattering diffusometry. Biomicrofluidics. 2016;10(5):054107. doi: 10.1063/1.4962992.

- [25] Meléndrez MF, Cárdenas G, Arbiol J. Synthesis and characterization of gallium colloidal nanoparticles. J Colloid Interface Sci. 2010;346(2):279–28.
- [26] Rauwel P, Küünal S, Ferdov S, Rauwel E. A review on the green synthesis of silver nanoparticles and their morphologies studied via TEM. Adv Mater Sci Eng. 2015;2015:682749.
- [27] Hedberg J, Skoglund S, Karlsson ME, Wold S, Odnevall Wallinder I, Hedberg Y. Sequential studies of silver released from silver nanoparticles in aqueous media simulating sweat, laundry detergent solutions and surface water. Env Sci Technol. 2014;48(13):7314–22. doi: 10.1021/es500234y. Epub 2014 Jun 19.
- [28] Rai MK, Deshmukh SD, Ingle AP, Gade AK. Silver nanoparticles: the powerful nanoweapon against multidrug-resistant bacteria. J Appl Microbiol. 2012;112:841–52.
- [29] Nel AE, M\u00e4dler L, Velegol D, Xia T, Hoek EMV, Somasundaran P, et al. Understanding biophysicochemical interactions at the nano-bio interface. Nat Mater. 2009;8:543-57.
- [30] Bilal M, Rasheed T, Iqbal HMN, Li C, Hu H, Zhang X. Development of silver nanoparticles loaded chitosan-alginate constructs with biomedical potentialities. Int J Biol Macromol. 2017;105(1):393–400.
- [31] Blagoi YuP, Galkin VL, Gladchenko GO, Kornilova SV, Sorokin VA, Shkorbatov AG. Metal complexes of nucleic acids in solutions. Naukova Dumka. 1991; p. 272.